

Innovative strategies in biological control

Stefan Vidal

Georg-August-University, Department of Crop Sciences, Goettingen

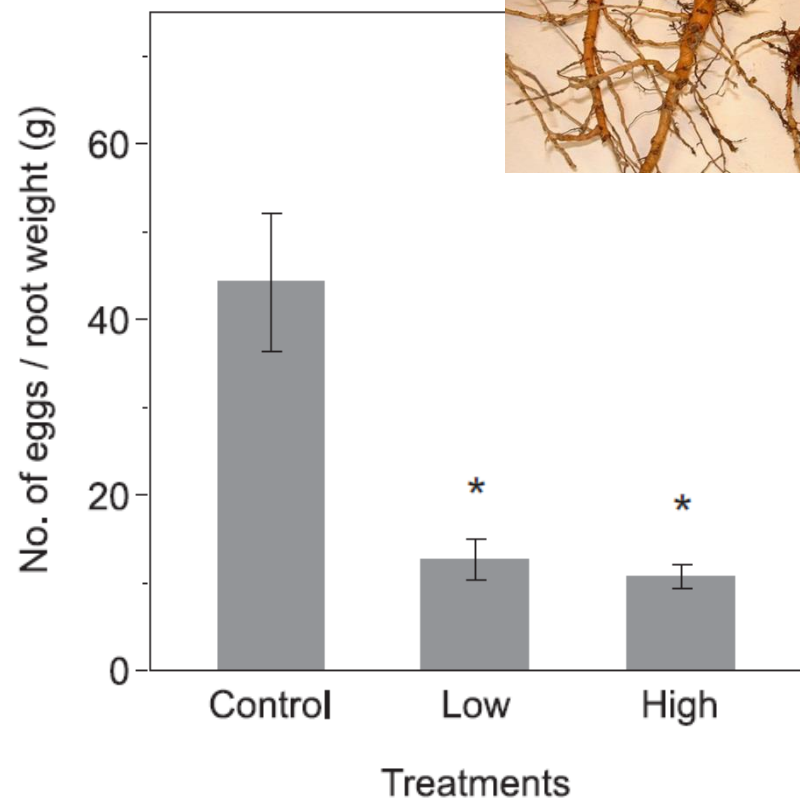
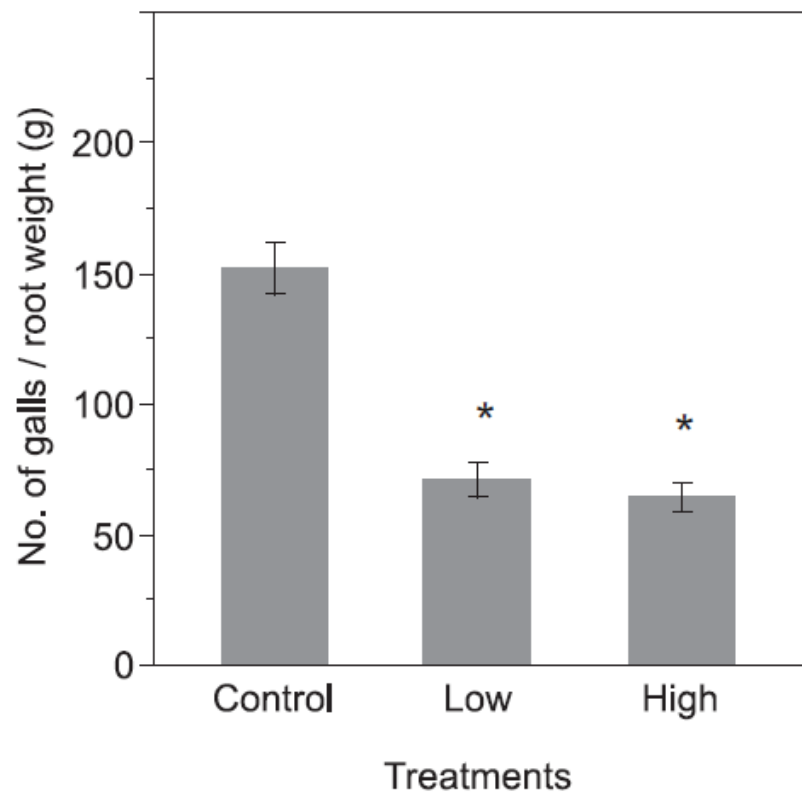
- Political background for implementation of biological control options better than ever
- Public awareness important
- New techniques/methodologies available
- CGR for BCAs at 15 % annually

Endophytic fungi as biocontrol agents

What do we know about endophytic fungi?

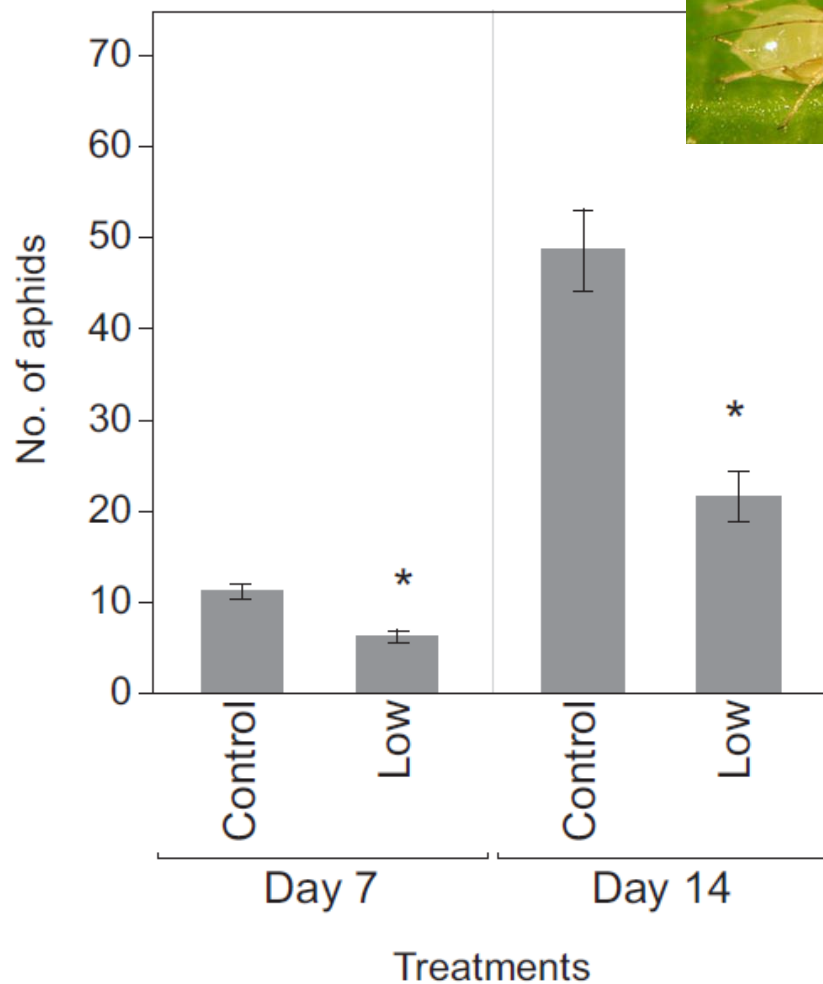
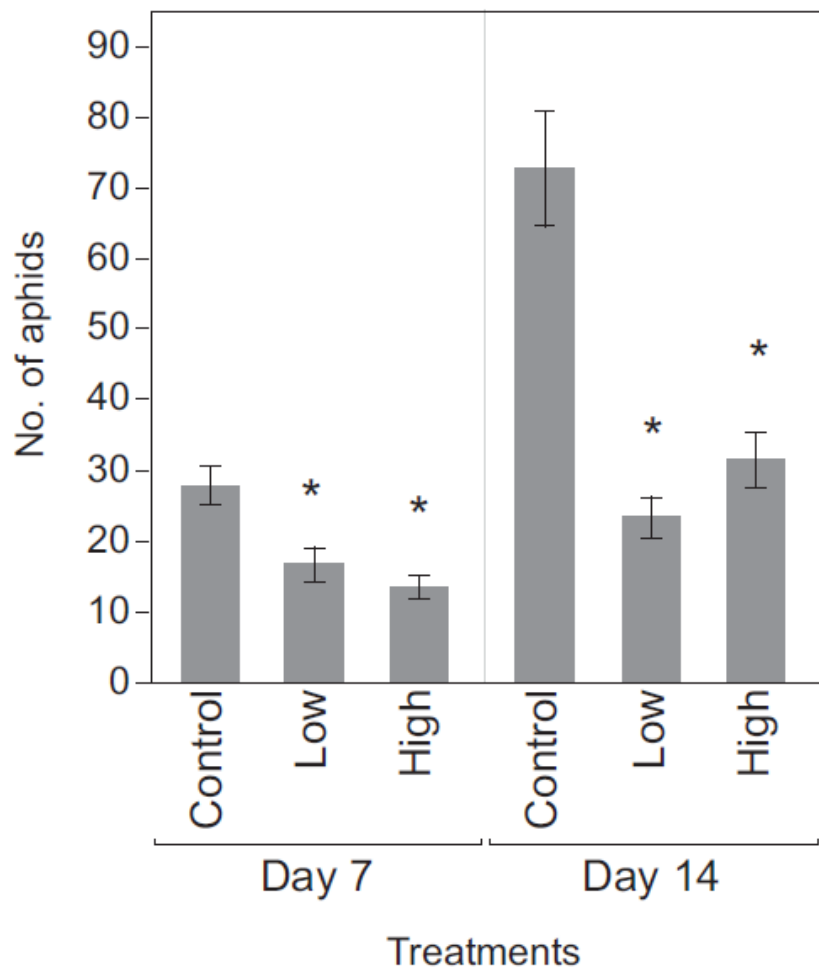
- Do not cause visible symptoms when colonizing plants
- Found in virtually all plant species (and algae, and)
- Producers of an extraordinary array of metabolites
- Plant growth promoters; antagonists of plant pathogens (and)

The fungal endophyte *Chaetomium globosum* negatively affects root knot nematodes on cotton

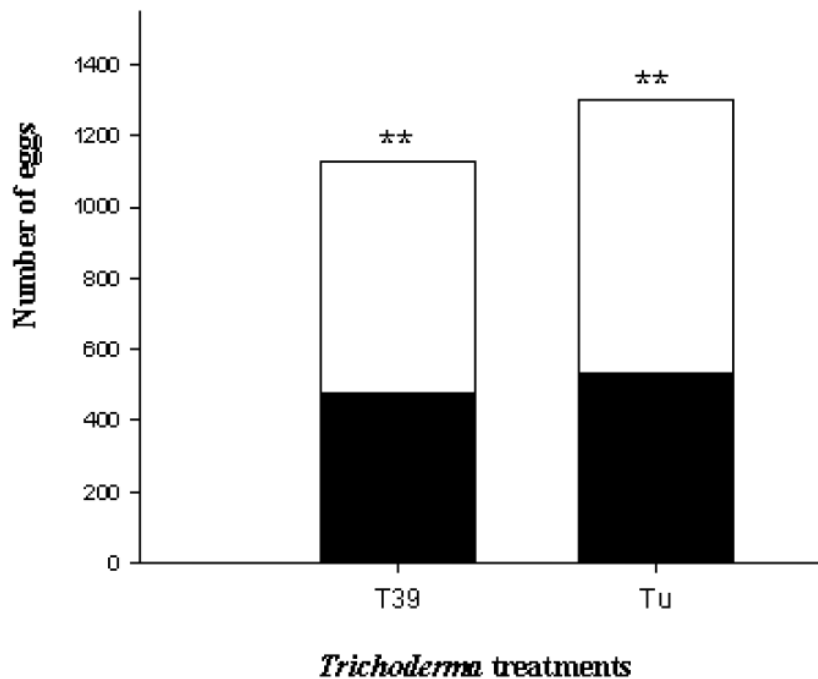




Low = 10^6 ; High = 10^7

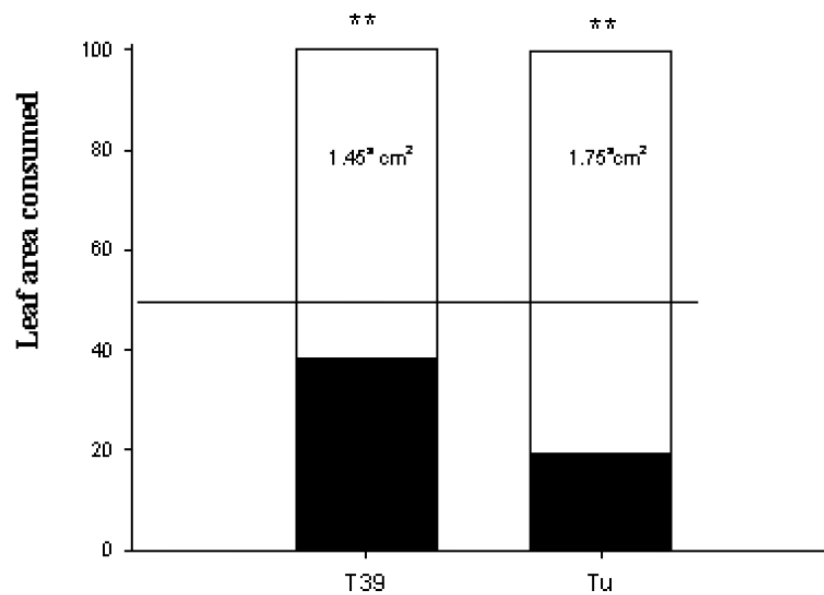
C. globosum also negatively affects cotton aphid reproduction



Oviposition and feeding of Diamondback moth larva on *Trichoderma* spp. inoculated cabbage plants in dual-choice assays



 Control
 Inoculated

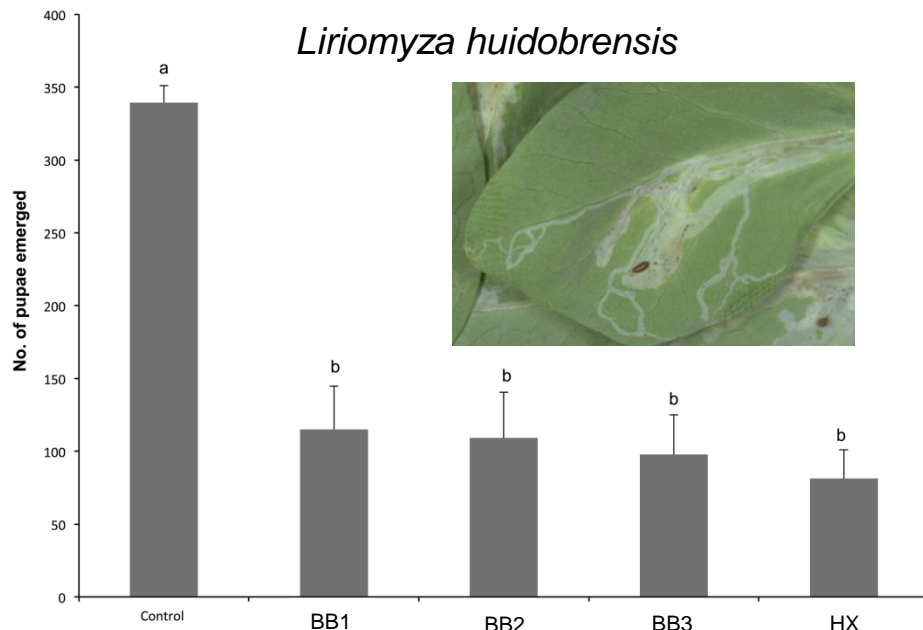


Virulence of 12 *B. bassiana* endophytic isolates/strains against third instar *H. armigera* larvae fed on leaves of inoculated *V. faba* plants

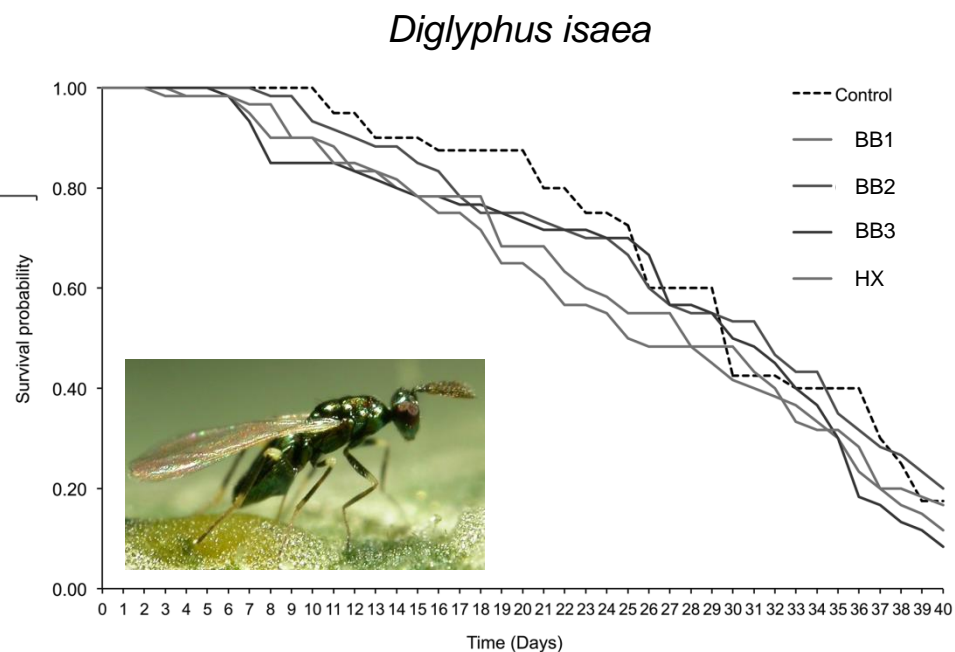
Treatment <i>B. bassiana</i> isolate	Parameter sampled \pm SE		
	Mortality (%)	Mycosis (%)	Survival time (days)
ATP01	70.00 \pm 0.11 ab ^a	00.00 \pm 0.00 b	10.36 \pm 0.82 b
ATP02	85.00 \pm 0.08 a	100.00 \pm 0.00 a	6.41 \pm 0.58 a
ATP03	10.00 \pm 0.07 cd	00.00 \pm 0.00 b	23.5 \pm 1.50 cd
ATP04	30.00 \pm 0.11 bcd	16.76 \pm 0.17 b	21.33 \pm 0.67 cd
ATP05	40.00 \pm 0.11 abcd	37.50 \pm 0.18 b	20.63 \pm 0.59 cd
Bb03032	55.00 \pm 0.11 abc	54.55 \pm 0.16 a	18.64 \pm 0.64 c
EABb04/01-Tip	45.00 \pm 0.11 abcd	66.67 \pm 0.17 a	19.11 \pm 0.63 c
Bb64	40.00 \pm 0.11 abcd	50.00 \pm 0.19 ab	20.25 \pm 0.59 c
Bb135	25.00 \pm 0.10 bcd	40.00 \pm 0.25 ab	20.80 \pm 0.66 cd
Bb1022	30.00 \pm 0.11 bcd	00.00 \pm 0.00 b	21.00 \pm 0.97 cd
Bb1025	35.00 \pm 0.11 bcd	28.57 \pm 0.18 b	20.29 \pm 0.67 c
Naturalis [®] (strain ATCC74040-based bioinsecticide)	25.00 \pm 0.10 bcd	22.22 \pm 0.15 b	21.11 \pm 0.68 cd
Control	00.00 \pm 0.00 d	00.00 \pm 0.00 b	24.60 \pm 0.83 d

^aMeans (\pm SE) followed by the same letter within a column are not significantly different at $P < 0.05$ (Tukey's HSD test with Bonferroni correction for multiple testing).

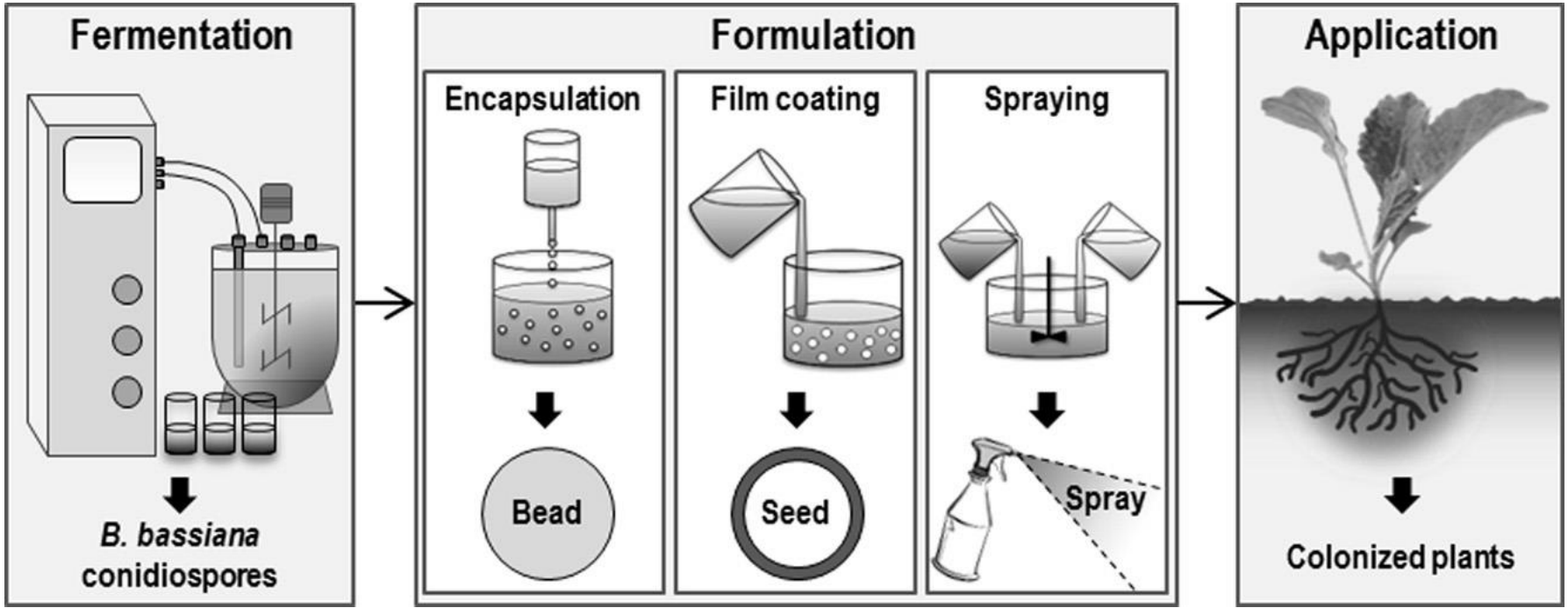
Endophyte colonization of *Vicia faba* by entomopathogenic fungi Isolates and life history of leafminer and parasitoid



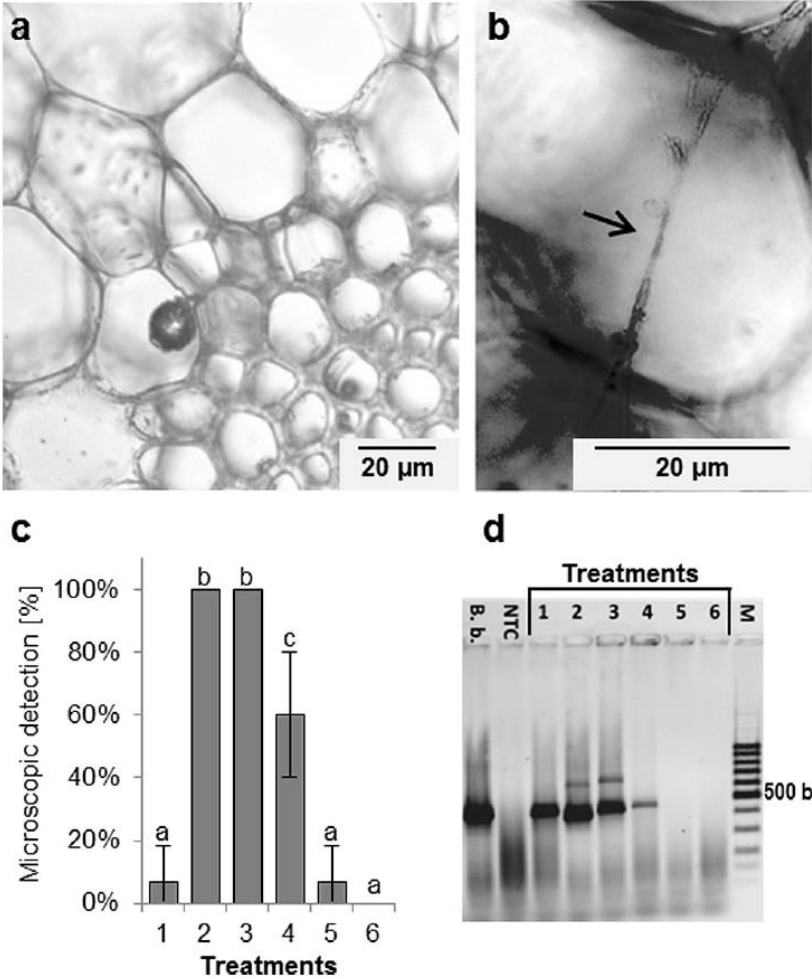
Beauveria bassiana (**BB**)
Hypocrea lixii (**HX**)



Different strategies to enhance endophytic colonization of entomopathogenic fungi in plant tissues



Influence of compositions from liquid formulations on endophytic colonization of oilseed rape leaves with *B. bassiana* after 14 days



Treatments:

1: water, *B. bassiana* (BB) - 2: Triton X-114 (Tri), sugar beet molasses (M), Titanium oxide (Td), BB
 3: TRI, M, BB - 4: Tri, TD, BB - 5: M, TD, BB - 6: TRI, M, TD

Pros and cons of using endophytic fungi

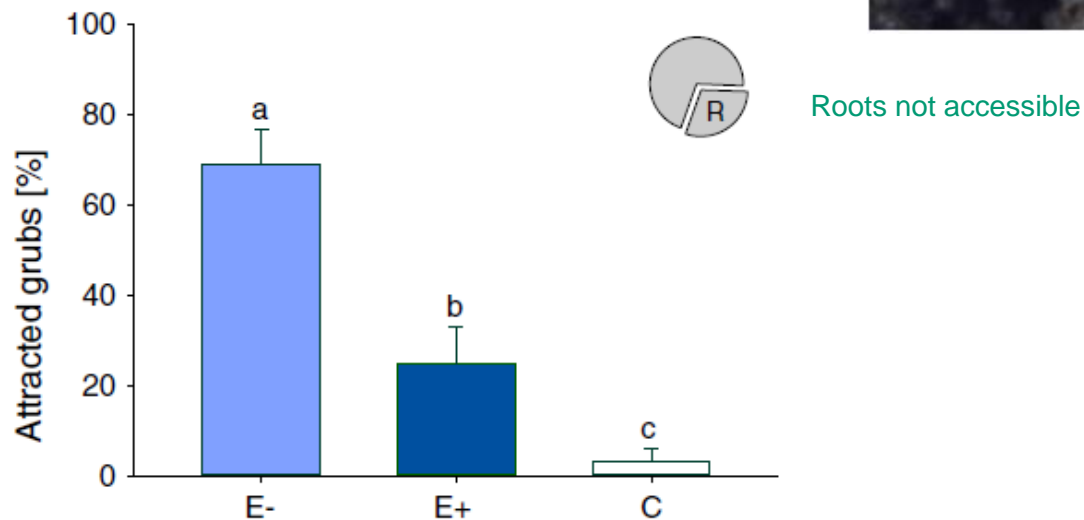
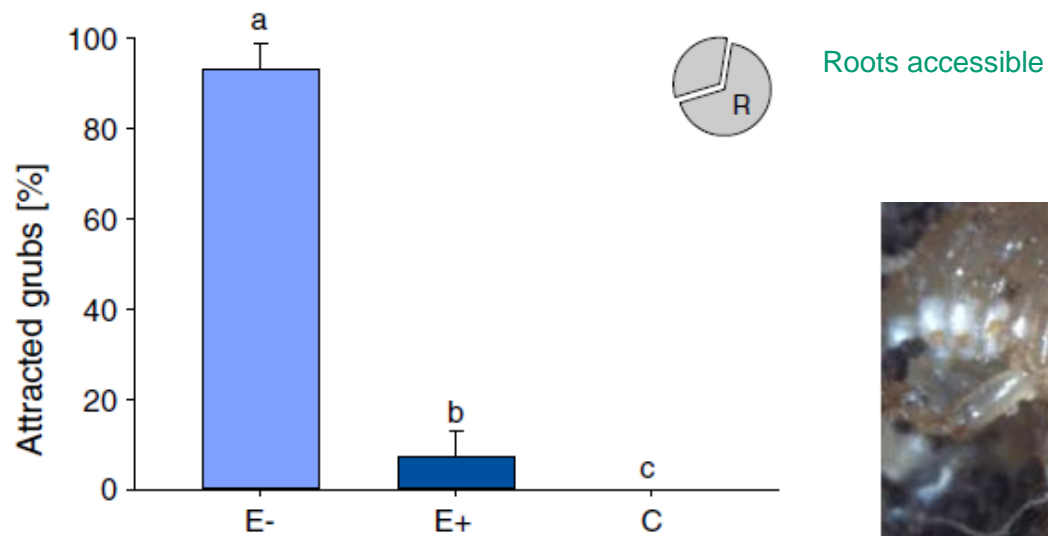
- + Specific isolates may target specific pest species
- + Entomopathogenic fungi are also working as endophytes
- + Mass production possible; costs therefore reasonable
- + Combination with other BCAs possible
- Colonization of all plant tissues not a trivial task
- Metabolite production in planta of concern
- Formulation issues not yet properly addressed

Use of multitrophic interactions

What do we know about multitrophic interactions?

- HIPVs produced by plants upon attack by herbivore pests
- Many of these HIPVs herbivore species specific
- Know to increase parasitism rates
- Cultivars differ in HIPV bouquets

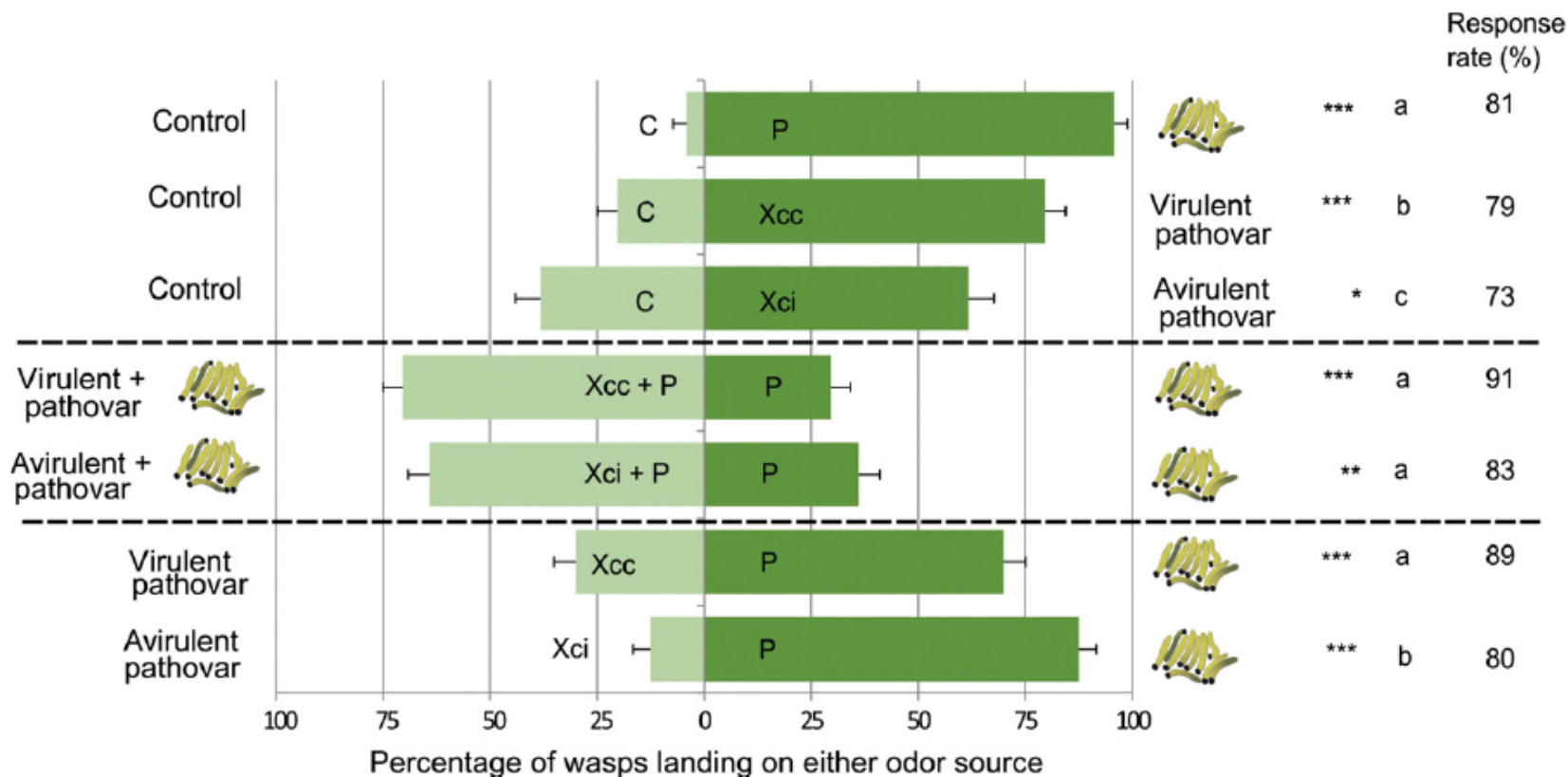
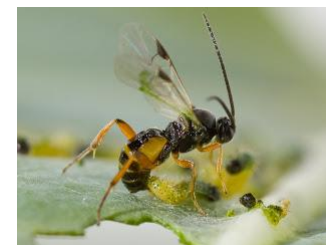
Response of *Costelytra zealandica* grubs to grass root volatiles in a four-arm olfactometer



Response rate of *Cotesia glomerata* in wind tunnel to *P. rapae* larvae on healthy and bacterium inoculated plant



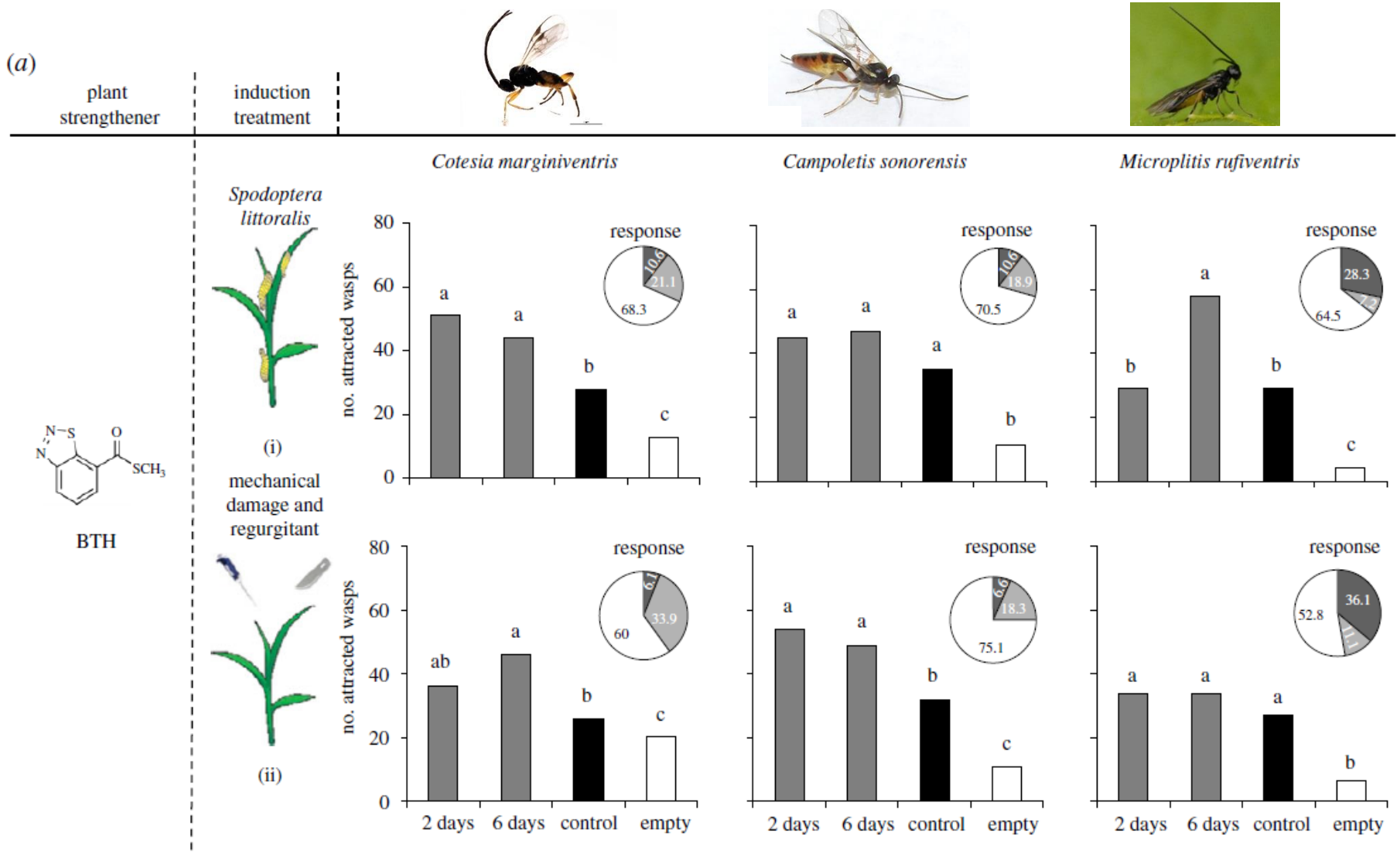
Xanthomonas campestris pv. campestris



Key compounds involved in stemborer control emitted by intercrop and trap plants in push-pull cropping systems

Pull semiochemistry					
Structure	Compound	Source	Method for ID	Target	Activity
	Octanal	Maize (<i>Zea mays</i>)	GC-EAG, GC-MS, co-elution with standards	Stemborer moths (adults)	Host cue attractant
	Nonanal	Napier grass			
	Naphthalene	Napier grass (<i>Pennisetum purpureum</i>)			
	4-allylanisole				
	Eugenol				
	Linalool				
Push semiochemistry					
*†	(<i>E</i>)-Ocimene	<i>Melinis minutiflora</i>	GC-EAG, GC-MS, co-elution with standards	Stemborer moths * <i>Cotesia sesamiae</i>	Repellent Attractant
*†	(<i>E</i>)-4,8-dimethyl-1,3,7-nonatriene	† <i>Desmodium</i> spp.			
	Humulene				
	β-Caryophyllene				
	α-Terpinolene				
†	α-Cedrene				

Enhancing the presence and efficacy of native biological control agents by applying plant strengtheners



BTH: benzo-(1,2,3)-thiadiazole-7-carbothioic acid S-methyl ester

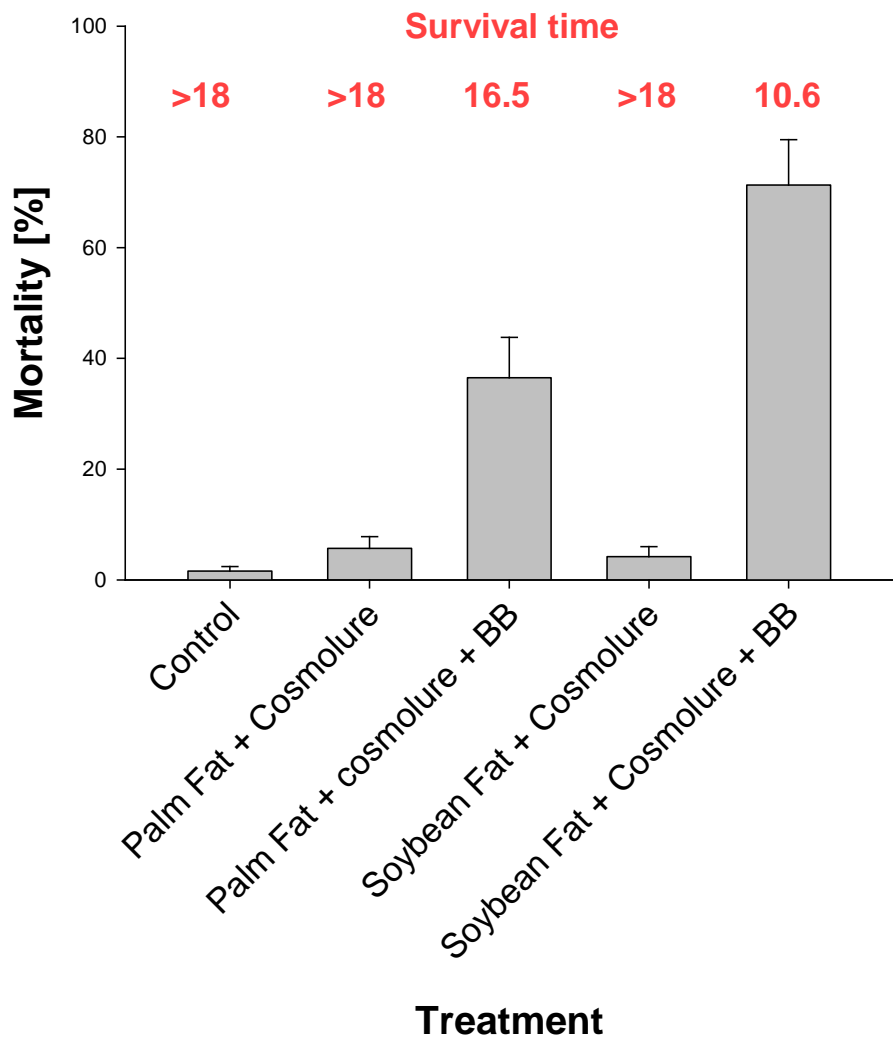
- + Compounds specifically attract specific parasitoids or predators
- + Non-target effects minimal
- Effects on pest population levels not clear
- Application costs a challenge
- Plant breeders not willing to step in

**Enhancing the efficacy of native
biological control agents by
combining different agents**

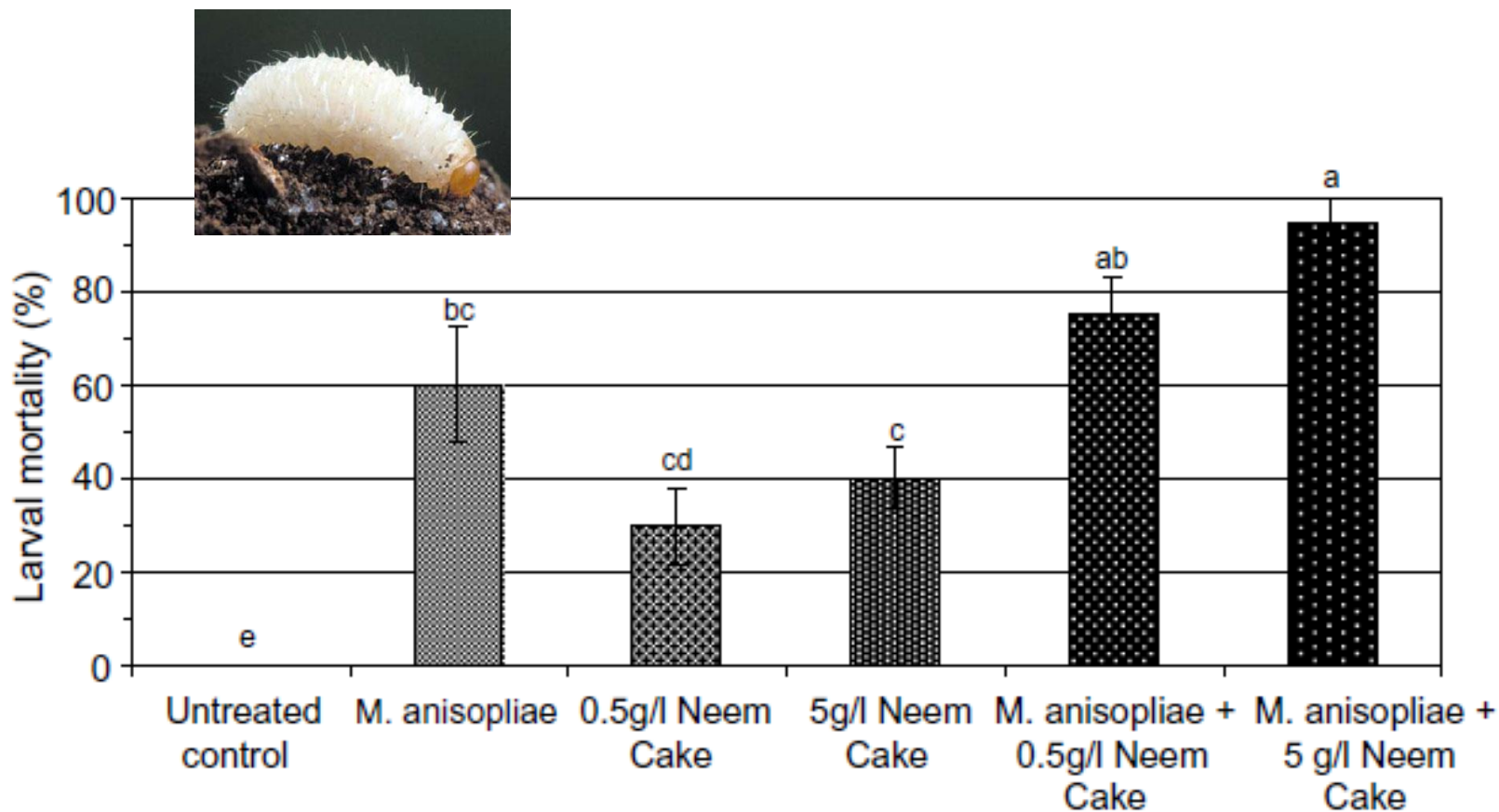
Combined use of a pheromone and an entomopathogenic fungus for banana weevil control



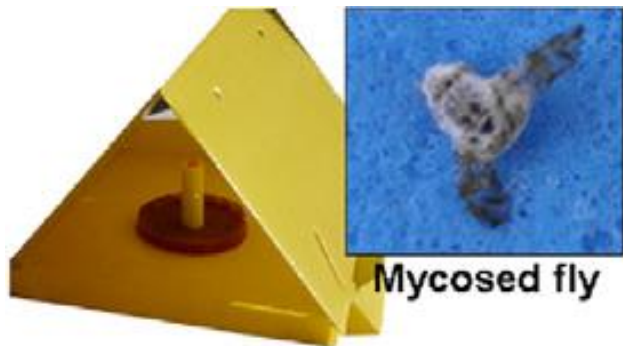
Cosmopolites sordidus



Mortality of BVW larvae in potted *Euonymus* plants treated with combinations of Neem cake or *Metarhizium* sp.



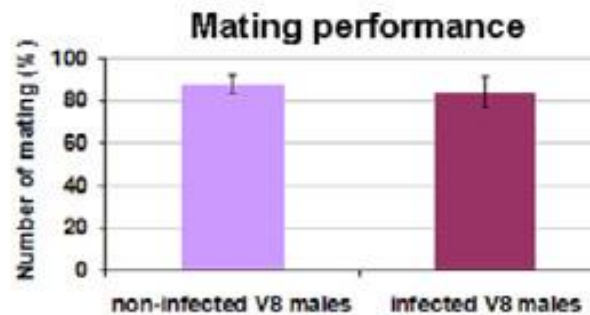
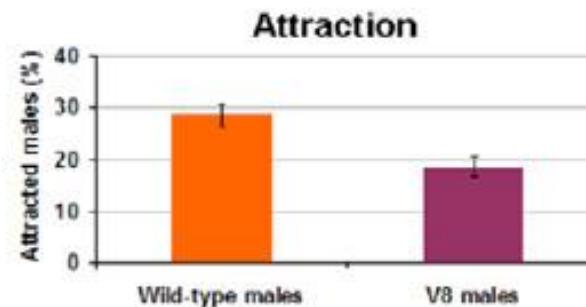
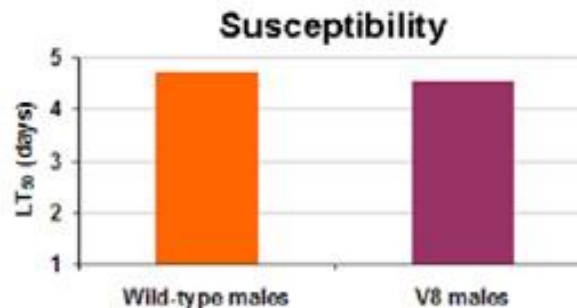
Combination of an attractant contaminant device containing an entomopathogenic fungus for Medfly control using sterile males



Attractant Contaminant Device



C. capitata V8 strain



- + Synergistic or additive effects possible
- + Non-target effects minimal
- Problems when it comes to registration
- Application costs too high (?)
- Potential combinations yet only marginally explored

Development of an “attract and kill” strategy for soil dwelling pests

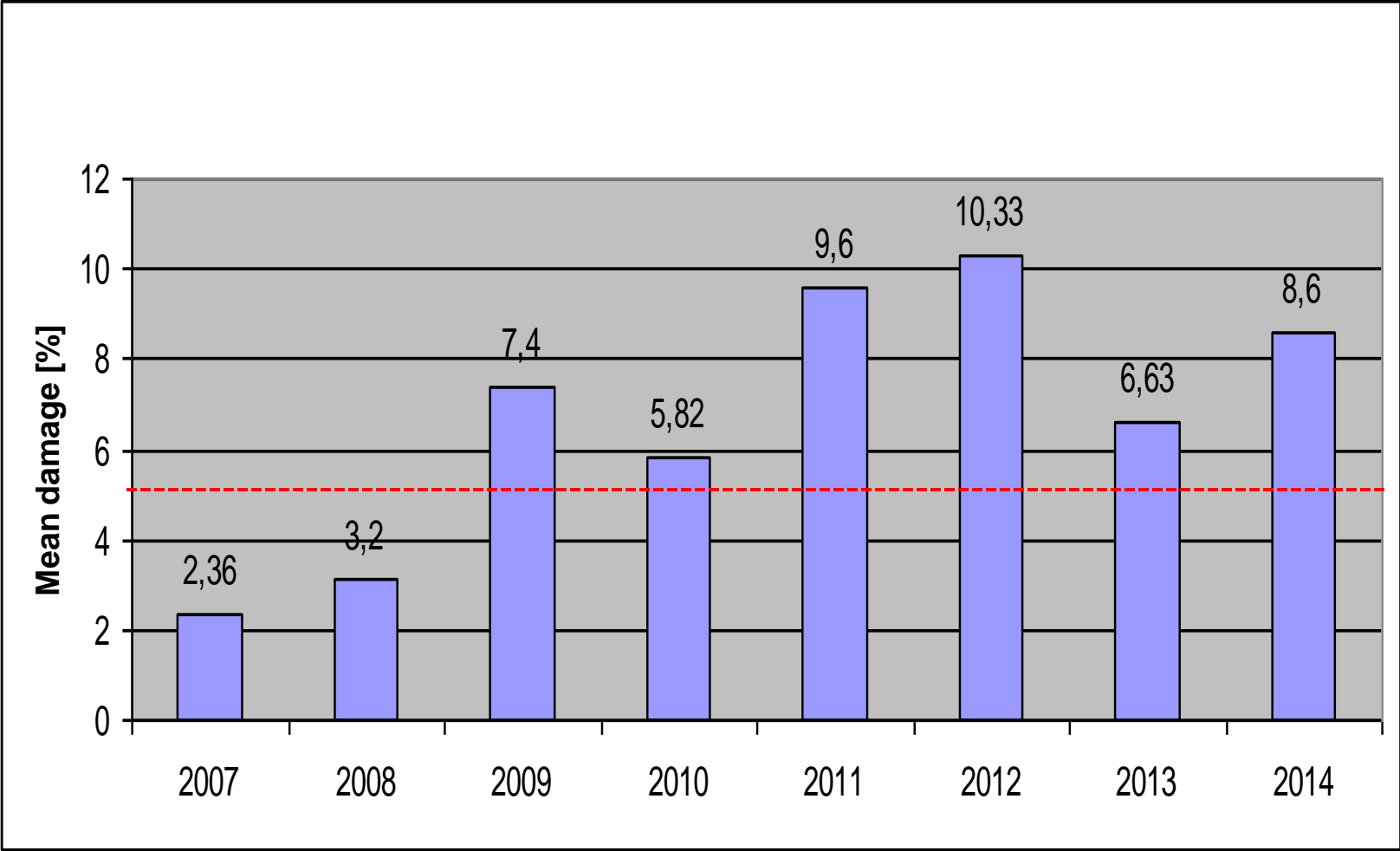
Most important common targets for new biocontrol products identified in arable, vegetable, and perennial crops in European

Pest	Crop(s)	Control solution	Available
Wireworms	Potato, maize	Entomopathogenic fungi	No - <i>yes</i>
Pollen beetle	Oilseed rape	Entomopathogenic fungi; entomopathogenic nematodes	No
Weevils	Oilseed rape	Parasitoids	No
Flea beetles	Oilseed rape	Entomopathogenic nematodes	No
Root flies	Brassicas	Beneficials	(Yes)
<i>Drosophila suzukii</i>	Stone & soft fruits	None	None

Damage on potato tubers caused by wireworms



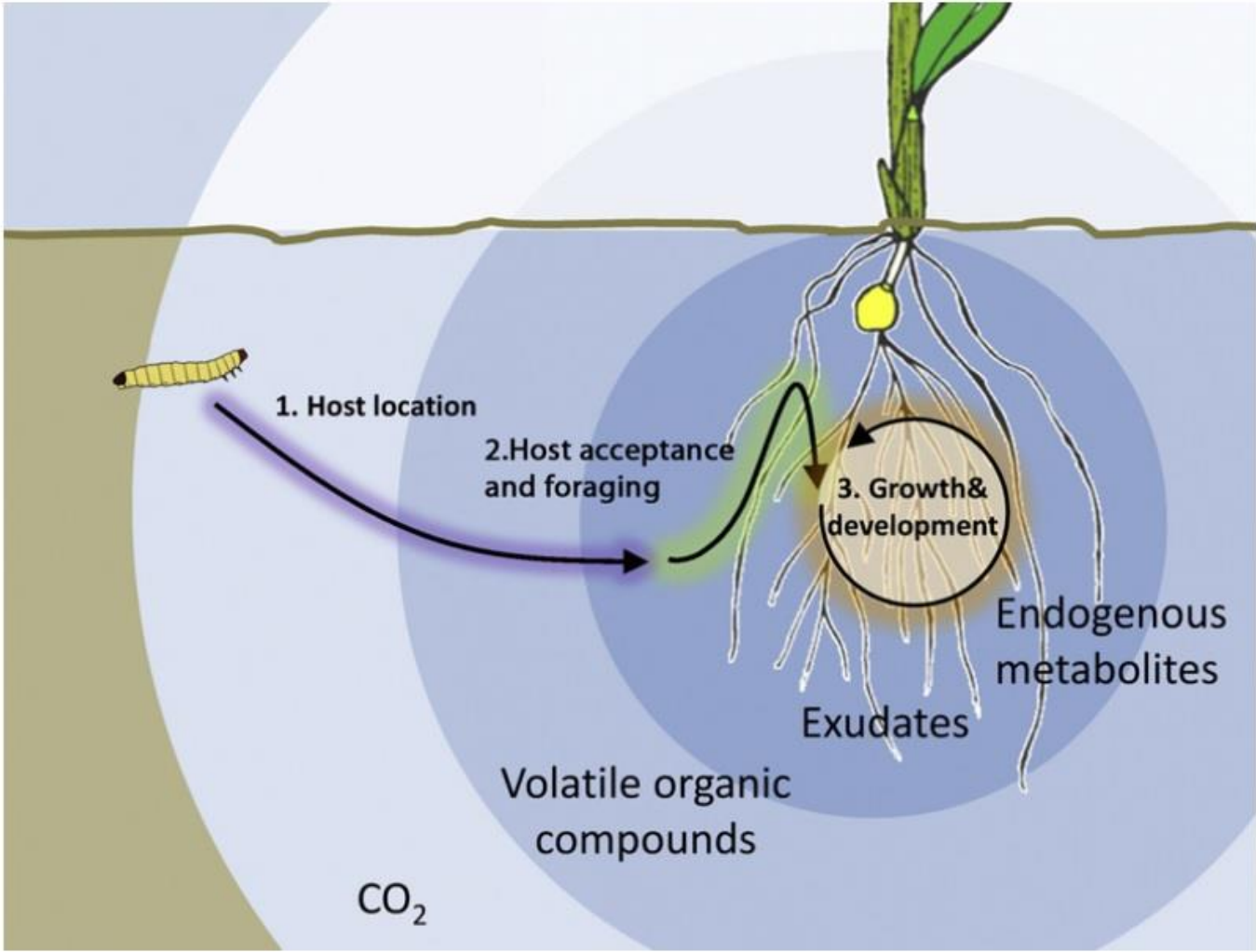
Mean wireworm damage in organic potato fields in Lower Saxony and Bavaria



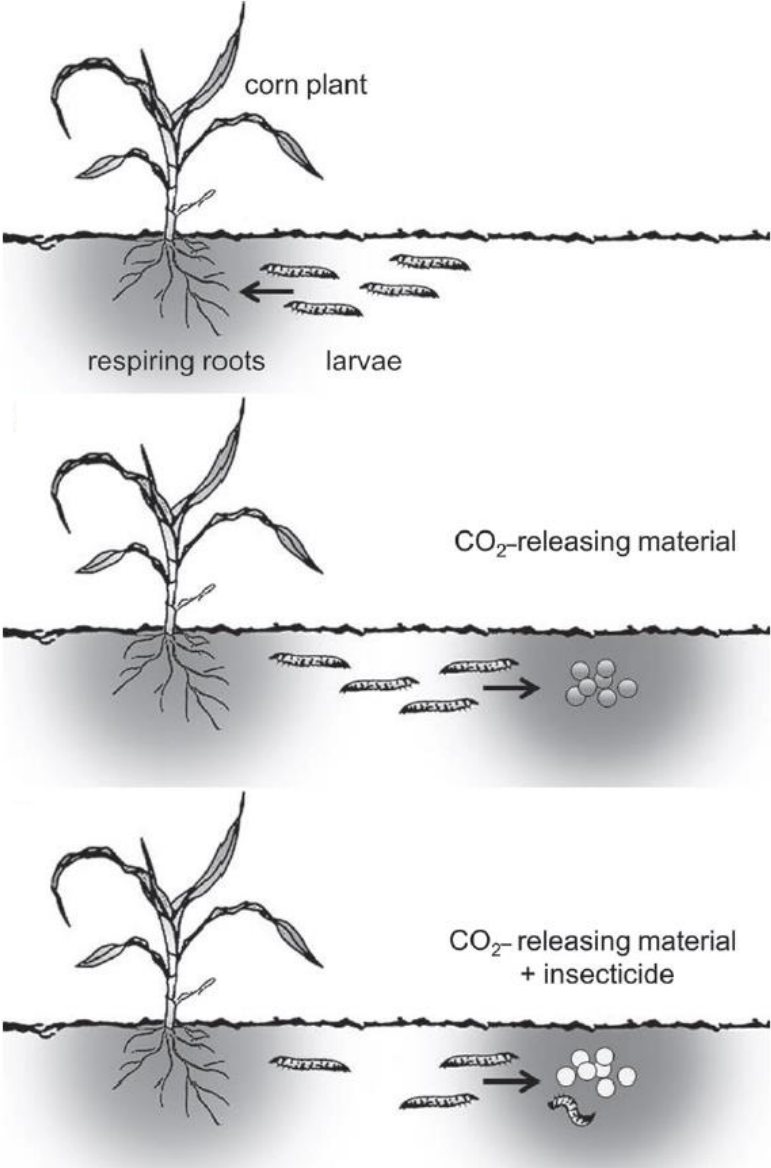
The problem

- Increasing wireworm damage reported in different crops (arable, vegetables, greenhouse)
- Control of wireworm species difficult because of
 - “Old” insecticides banned (i.e. Lindan)
 - Phasing out of efficient a.i.s (Neonicotinoids, Fipronil (?))
 - Application techniques mostly not suitable
 - Patchy distribution

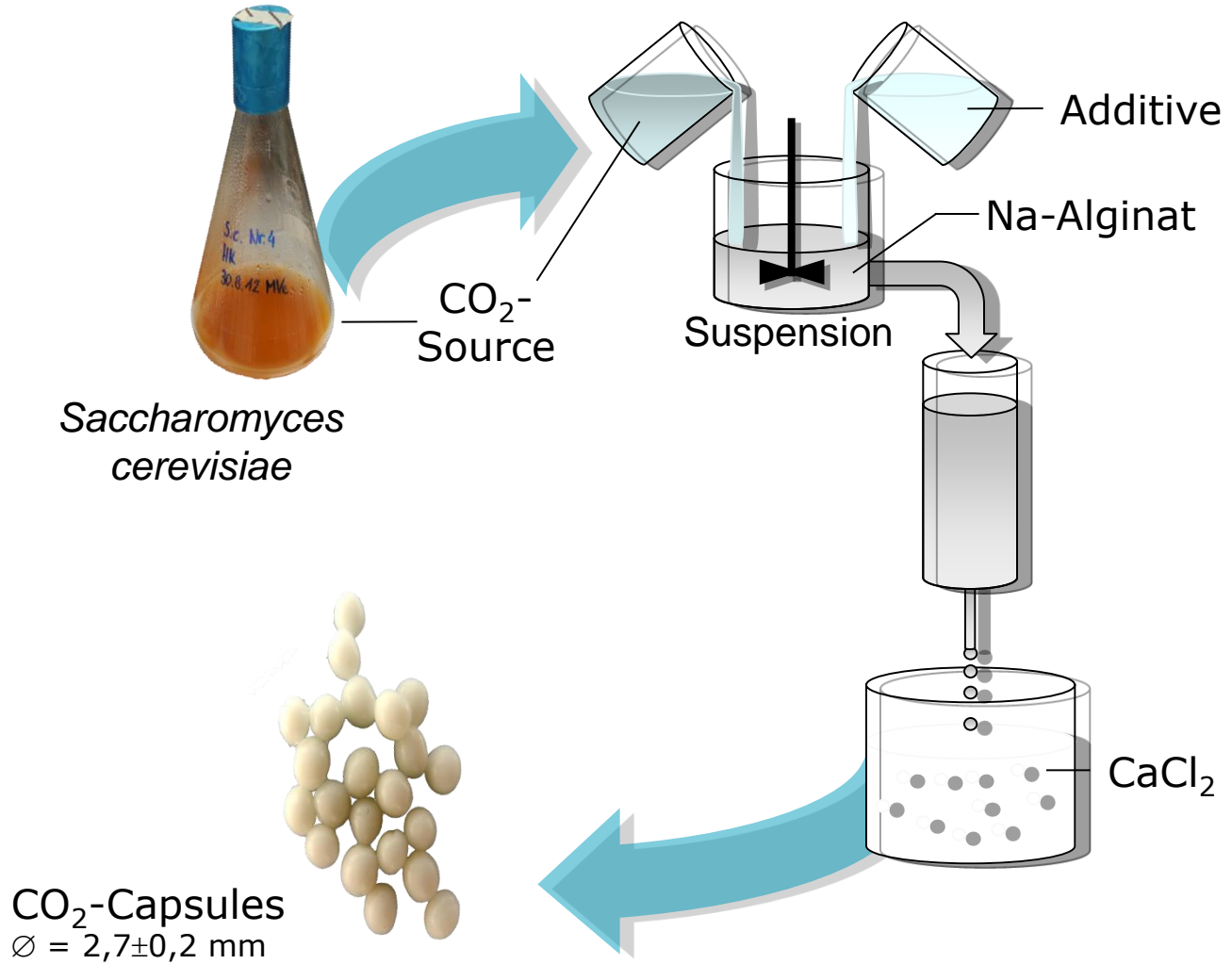
Orientation cues of (most) soil dwelling larvae



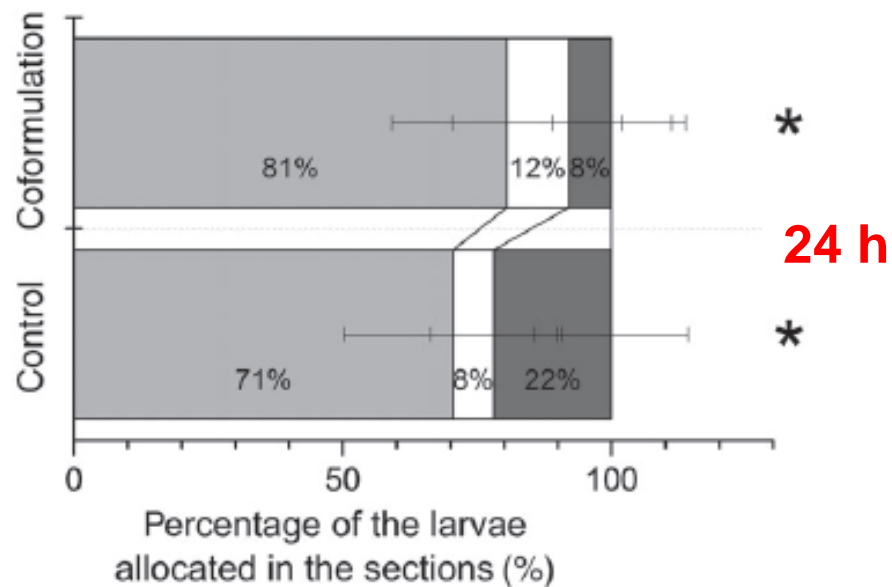
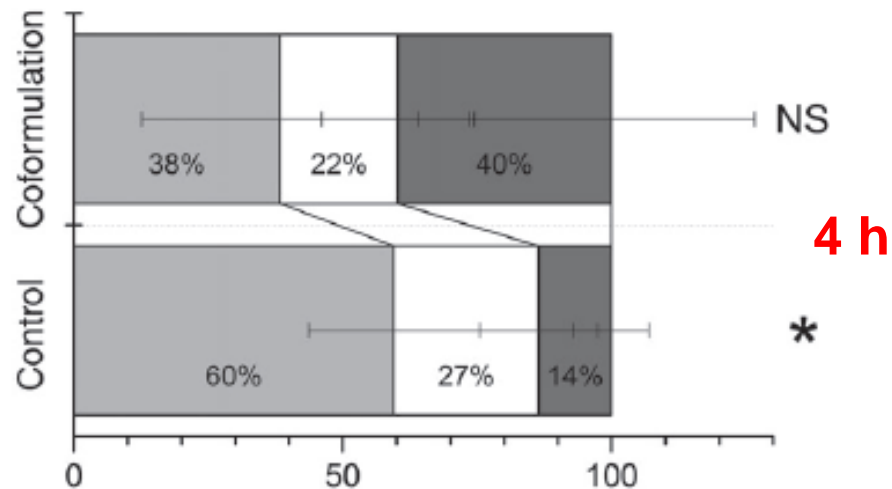
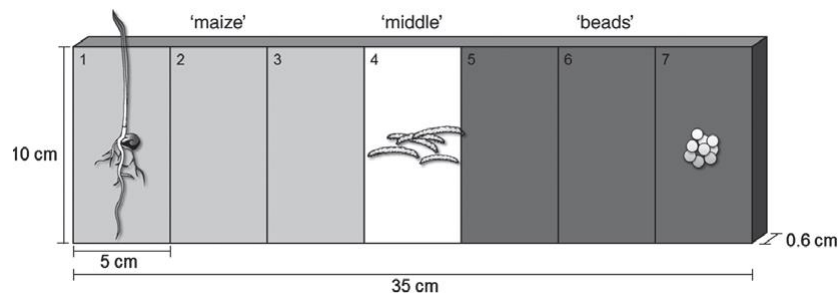
Orientation of soil-dwelling larvae towards CO₂ gradients of growing maize plants



Production of CO₂ emitting capsules



Orientation of western corn rootworm larvae when exposed to maize roots or CO₂ releasing compounds



Addition of a kill-component

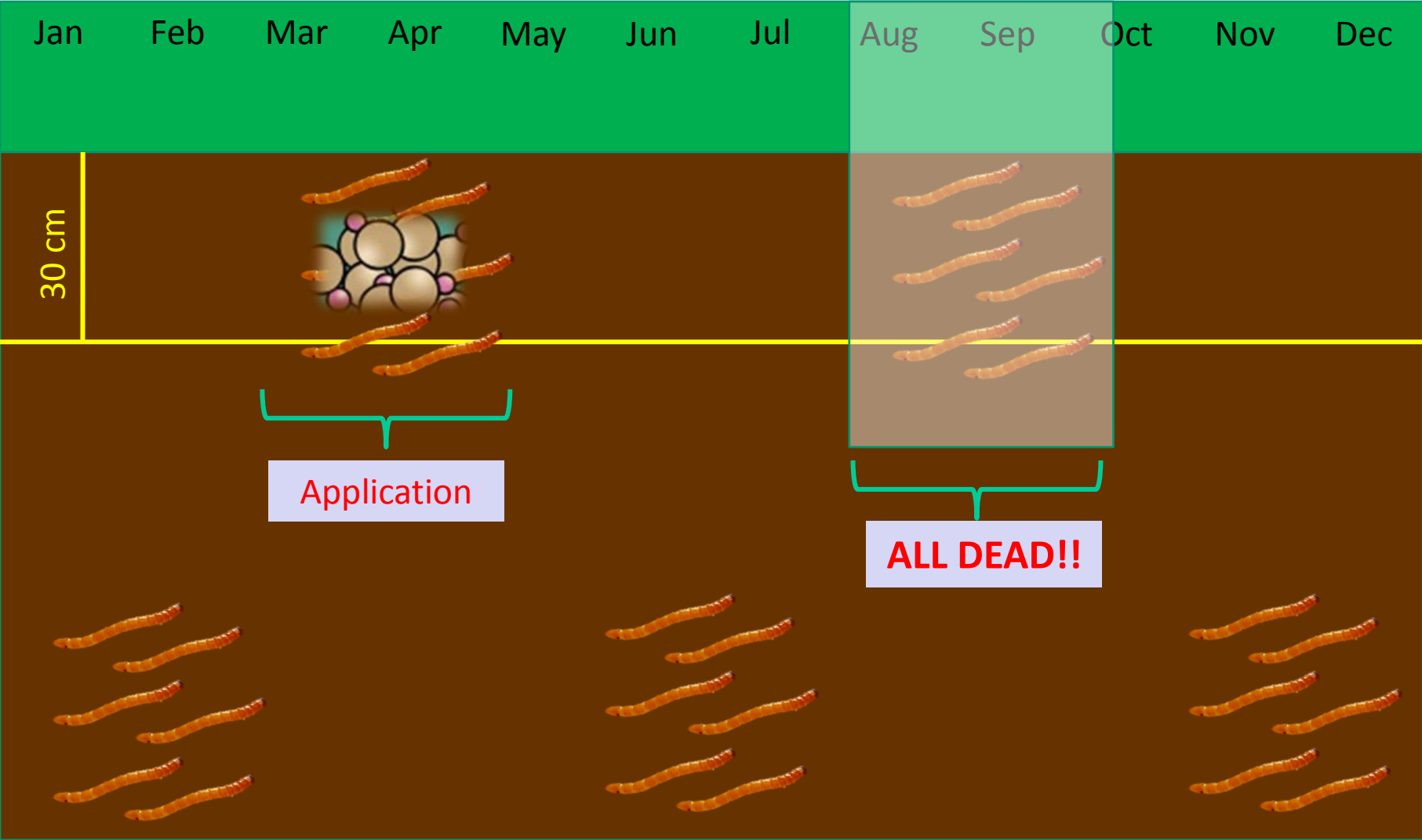
- Insecticides (*Force 1.5G*)
- Spinosad
- Neem
- Entomopathogenic fungi

(*Metarhizium brunneum* [ART 2825])



~ 4 weeks

Vertical movement of wireworms and window of vulnerability



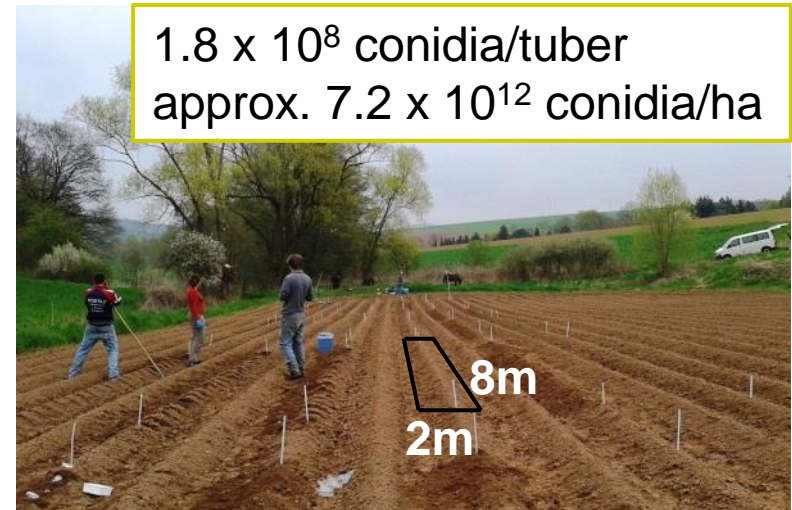
Experimental set-up

- Fields in different federal states (2013-15)
- Conventional, organic farming
- Randomized block design
 - 6 to 8 reps (EPPO guideline)
 - Plot: 1 potato row (8 m length)
 - Untreated buffer rows

Treatments

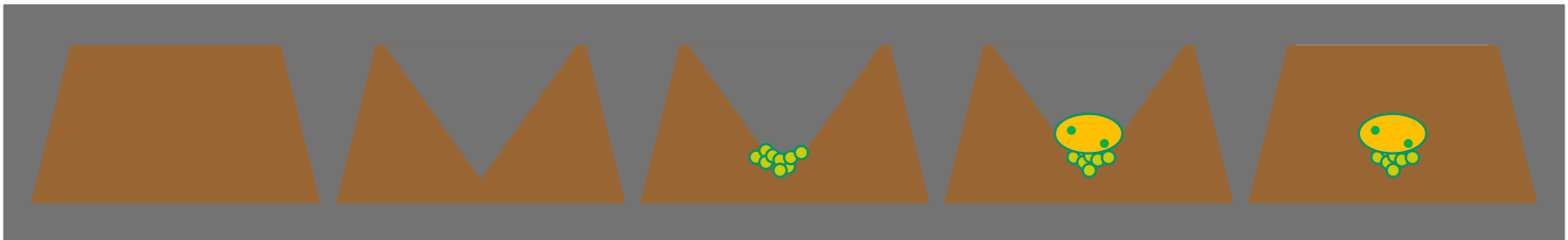
– Control

- *M. brunneum* - suspension
- *M. brunneum* - beads
- CO₂-beads
- Attract + Kill – beads
- Firponil (Goldor Bait®)



• Applications

- Spot (Beneath, between)
- Band (Beneath)



Every beginning is difficult: Spoon-application-technology



Field tests using co-formulated A&K compounds



AK-capsules

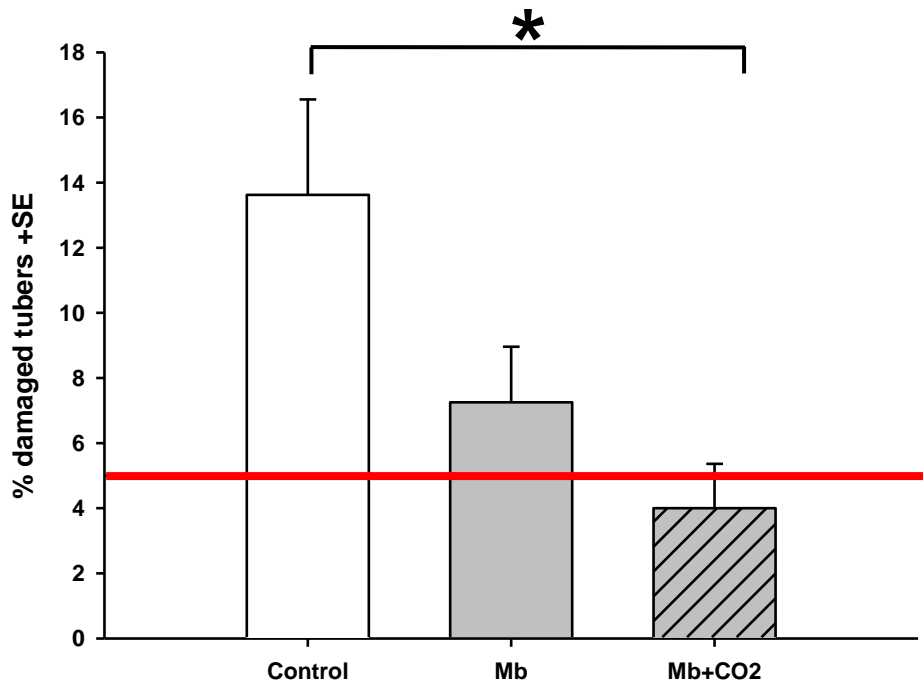


AK-granules

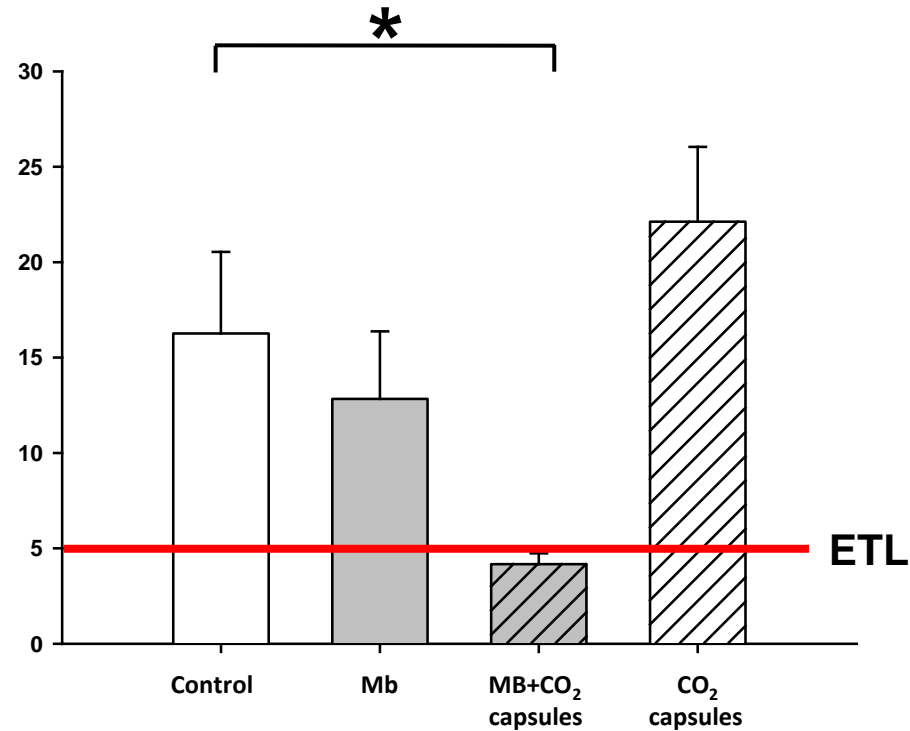


Damage assessment on potato tubers: *Metarhizium brunneum* as the killing agent

Waake 2014



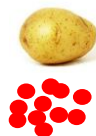
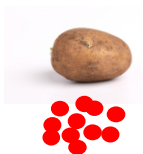
HamelIn 2014



Kruskal-Wallis: $H_{2,24} = 7.09$ $P < 0.05$

Treatments

Kruskal-Wallis: $H_{3,31} = 9.81$ $P < 0.05$





neu

Innovativer Pflanzenschutz
für den Kartoffelanbau!

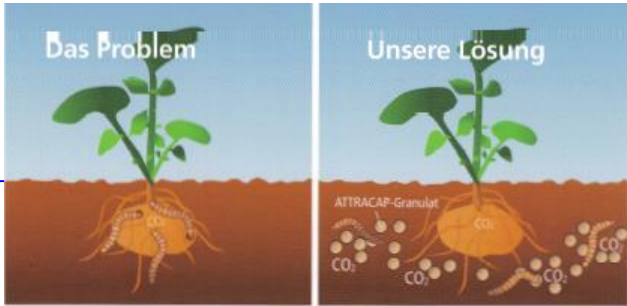
**Biologische
Drahtwurm-
Bekämpfung**



ATTRACAP



Made in Germany



ATTRACAP

ATTRACAP von **BIOCARE** ist ein innovatives biologisches Insektizid zur Bekämpfung von Drahtwürmern. Es ist zugelassen nach Artikel 53 für Notfallsituationen (01.03.2016–28.06.2016). Im Kartoffelanbau haben wirtschaftliche Verluste durch Drahtwurmbefall erheblich zugenommen. Drahtwürmer, die Larven des Schnellkäfers, fressen tiefe Löcher in die Kartoffeln, die dadurch teilweise nicht mehr vermarktet werden können.

Die Wurzeln der Kartoffeln geben CO₂ ab und locken damit die Drahtwürmer an. Die **Attract-and-Kill-Strategie** von **ATTRACAP** nutzt dies. Das Produkt erzeugt über ca. 4 Wochen CO₂, wodurch die Drahtwürmer angelockt werden. Neben der CO₂-Quelle befindet sich im Granulat der natürlich vorkommende, insektenabtötende Pilz *Metarhizium brunneum*. Dieser wächst aus dem Granulat aus. Durch den Kontakt mit den ausgewachsenen Sporen infizieren sich die Larven und sterben nach einigen Tagen ab.

- Anwendung**
- Befallsstärke:** Niedriger bis mittlerer Befall
 - Wirkstoff:** *Metarhizium brunneum*
 - Ausbringung:** 30 kg je Hektar
 - Ausbringungsmethode:** Granulatstreuer (JKI-geprüft)

Pflanzenschutzmittel vorsichtig verwenden. Vor Verwendung stets Etikett und Produktinformationen lesen!

BIOCARE-Forschungsförderung durch:



BIOCARE Gesellschaft für Biologische Schutzmittel mbH
Dorfstr. 4 ■ D-37574 Einbeck ■ Tel. (0 55 62) 95 05 78-0
Fax (0 55 62) 95 05 78-9 ■ E-Mail: info@biocare.de ■ www.biocare.de

1107/2009 EG Article 53
Emergency situations in PP

Summary of efficacy data from independent farmer field trials 2016

Landwirt	Region	Befall im Vorjahr	Bekämpfung im Vorjahr	Kultur im Vorjahr	Bodentyp	Beregnung	Anzahl befallener Knollen / 100 Knollen		WG %	Bemerkung
							unbehandelte Fläche	behandelte Fläche		
1	Vorderpfalz / Rheinland-Pfalz	schwach	ja	Zuckerrüben, Sommergerste	sandiger Lehm	ja	0	0	-	nicht bewertet
2	Vorderpfalz / Rheinland-Pfalz	schwach	ja	Sommerzwiebel	uL	ja	3	3	0	nicht bewertet, da unter dern ökonomischen Schadensschwelle
3	Rheinessen / Pfalz	schwach	ja	Kartoffel	sL	ja	40	10	75	
4	Rheinland-Pfalz	schwach	ja	Getreide	sL	ja	keine	0	-	nicht analysierbar, da keine Kontrolle vorhanden
5	Rheinland-Pfalz	mittel	nein	Karotten	IS	ja	23	11	52	
6	Rheinland-Pfalz	unbekannt	nein	Winterweizen	sL	ja (überschwemmt)	keine	3,5	-	nicht analysierbar, da keine Kontrolle vorhanden
7	Vorderpfalz / Rheinland-Pfalz	stark	nein	Zwiebeln	sL	ja	keine	6	-	nicht analysierbar, da keine Kontrolle vorhanden
8	Worms / Pfalz	nein	nein	Weizen	sL	ja	3	1	67	nicht bewertet, da unter dern ökonomischen Schadensschwelle
9*	Wittlich, Rheinland-Pfalz			Raps	sL	nein	20	0	100	
10	NRW	mittel	ja	Wintergerste	sL	ja	30	15	50	
10	NRW	mittel	ja	Wintergerste	sL	ja	30	4	87	

\emptyset	72,8
SD	21,8

Conclusions

- New technologies provide opportunities targeting organisms not yet controlled
- The potential for innovative formulations, combining different BCAs has not yet been fully exploited
- Artificial CO_2 -gradients are attractive for soil dwelling insects
- CO_2 -releasing capsules offer new opportunities to increase the efficacy of biocontrol agents
- The attract & Kill-strategy can be extended to other target organisms

Project funds for „A & K“ since 2012

